
Biotechnology Explorer™

Chromosome 16: PV92 PCR Informatics Kit

Catalog #166-2100EDU

explorer.bio-rad.com

Note: Kit contains temperature-sensitive reagents. Open immediately upon arrival and store components at -20°C or at 4°C as indicated.

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Student Manual

Introduction to PCR — The Polymerase Chain Reaction

You are about to perform a procedure known as PCR¹ to amplify a specific sequence of your own DNA in a test tube. You will be looking for a particular piece of DNA that is present in the genes of many, but not all, people. Analysis of the data generated in this laboratory will enable you to determine whether or not you carry this specific DNA sequence.

The **genome**, composed of DNA, is our hereditary code. This is the so-called blueprint that controls much of our appearance, behavior, and tendencies. **Molecular biology** is the study of genes and the molecular details that regulate the flow of genetic information from DNA to RNA to proteins, from generation to generation. **Biotechnology** uses this knowledge to manipulate organisms' (microbes, plants, or animals) DNA to help solve human problems.

Within the molecular framework of biology, DNA, RNA, and proteins are closely tied to each other. Because proteins and enzymes ultimately play such a critical role in the life process, scientists have spent many lifetimes studying proteins in an attempt to understand how they work. With this understanding, it was believed we could cure, prevent, and overcome disease and physical handicaps as well as explain exactly how and why organisms exist, propagate, and die. However, the complete answer to how and why does not lie solely in the knowledge of how enzymes function; we must learn how they are made. If each enzyme is different, then what controls these differences and what is the blueprint for this difference? That answer lies within our genome, or genetic code.

Thus, you may realize why researchers today, in an attempt to understand the mechanisms behind the various biological processes, study nucleic acids as well as proteins to get a complete picture. In the last 20 years, many advances in nucleic acid techniques have allowed researchers to study the roles that nucleic acids play in biology. It took the imagination and hard work of many scientists to reveal the answers to one of the most mysterious puzzles of life — understanding the mechanisms that control how DNA is translated into proteins within living cells.

Before Beginning This Lab, See If You Can Answer the Following Questions

How is DNA faithfully passed on from generation to generation? What causes genetic disease in some people but not others? How do scientists obtain DNA to study? What secrets can DNA tell us about our origins? What human problems can an understanding of DNA help us solve? Should we unlock the secrets held in this most basic building block of life?

PCR Set the Stage for a Scientific Revolution

In 1983, Kary Mullis² at Cetus Corporation developed the molecular biology technique that has since revolutionized genetic research. This technique, called the **polymerase chain reaction** (PCR), transformed molecular biology into a multidisciplinary research field within 5 years of its invention. Before PCR, the molecular biology techniques used to study DNA required such a high level of expertise that relatively few scientists could use them.

The objective of PCR is to produce a large amount of DNA in a test tube (in vitro), starting from only a trace amount. Technically speaking, this means the controlled enzymatic amplification of a DNA sequence, or gene, of interest. The template strands can be any form of DNA, such as genomic DNA. A researcher can use tiny amounts of genomic DNA

from a drop of blood, a single hair follicle, or a cheek cell, and make enough DNA to study. In theory, only a single template strand is needed to copy and generate millions of new identical DNA molecules. Prior to PCR, this would have been impossible. It is the ability to amplify the precise sequence of DNA of interest that is the true power of PCR.

PCR has made an impact on four main areas of genetic research: gene mapping; cloning; DNA sequencing; and gene detection. PCR is now used as a medical diagnostic tool to detect specific mutations that may cause genetic disease;³ in criminal investigations and courts of law to identify suspects,⁴ and in the sequencing of the human genome.⁵ Prior to PCR, the use of molecular biology techniques for therapeutic, forensic, pharmaceutical, agricultural, or medical diagnostic purposes was neither practical nor cost-effective. The development of PCR transformed molecular biology from a difficult science to one of the most accessible and widely used disciplines of **biotechnology**.

Two methods for DNA template preparation are provided in the manual. Your instructor will indicate which exercise to follow. Now, let's extract some of your own DNA.

Lesson 1 Cheek Cell DNA Template Preparation

To obtain DNA for use in the polymerase chain reaction (PCR) you will extract the DNA from your own living cells. It is interesting to note that DNA can be also extracted from mummies and fossilized dinosaur bones. In this lab activity, you will isolate DNA from epithelial cells that line the inside of your cheek. To do this, you will rinse your mouth with a saline (salt) solution, and collect the cells using a centrifuge. You will then boil the cells to rupture them and release the DNA they contain. To obtain pure DNA for PCR, you will use the following procedure:

The cheek cells are transferred to a micro test tube containing **InstaGene™ matrix**. This particulate matrix is made up of negatively charged, microscopic beads that chelate, or grab, metal ions out of solution. It traps metal ions, such as Mg^{2+} , which are required as catalysts or **cofactors** in enzymatic reactions. Your cheek cells will then be **lysed**, or ruptured, by heating to release all of their cellular constituents, including enzymes that were once contained in the cheek-cell lysosomes. Lysosomes are sacs in the cytoplasm that contain powerful enzymes, such as **DNases**, which are used by cells to digest the DNA of invading viruses. When you rupture the cells, these DNases can digest the released DNA. However, when the cells are lysed in the presence of the chelating beads, the cofactors are adsorbed and are not available to the enzymes. This virtually blocks enzymatic degradation of the extracted DNA so you can use it as the template in your PCR reaction.

You will first suspend your isolated cheek cells in the InstaGene matrix and incubate them at 56°C for 10 minutes. This preincubation step helps to soften plasma membranes and release clumps of cells from each other. The heat also inactivates enzymes, such as DNases, which can degrade the DNA template. After this 10 minute incubation period, place the cells in a boiling (100°C) water bath for 5 minutes. Boiling ruptures the cells and releases DNA from their nuclei. You will use the extracted genomic DNA as the target template for PCR amplification.

Lesson 1 Cheek Cell DNA Template Preparation (Lab Protocol)

Workstation Checklist

Materials and supplies required at the workstations prior to beginning this exercise are listed below.

Student Workstation	Quantity per Station	(✓)
1.5 ml micro test tubes	4	<input type="checkbox"/>
Screwcap tubes with 200 μ l InstaGene™ matrix	4	<input type="checkbox"/>
Foam micro test-tube holder	2	<input type="checkbox"/>
P-20 micropipet, 2–20 μ l	1	<input type="checkbox"/>
Pipet tips (filter type), 2–20 μ l	4	<input type="checkbox"/>
Permanent marker	1	<input type="checkbox"/>
Copy of Quick Guide or protocol	1	<input type="checkbox"/>
Waste container	1	<input type="checkbox"/>
Cups with 10 ml 0.9% saline	4	<input type="checkbox"/>

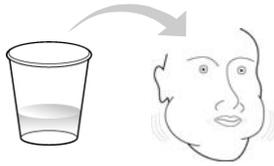
Instructor's Workstation	Quantity per Class	
P-20 micropipet, 2–20 μ l	1	<input type="checkbox"/>
P-200 micropipet, 20–200 μ l	1	<input type="checkbox"/>
Pipet tips (filter type), 2–20 μ l	1 box	<input type="checkbox"/>
Pipet tips (filter type), 20–200 μ l	1 box	<input type="checkbox"/>
Water baths (56 and 100°C)	1 each	<input type="checkbox"/>
Microcentrifuge	1	<input type="checkbox"/>
or mini centrifuge	4	<input type="checkbox"/>
Vortexer (optional)	1	<input type="checkbox"/>

Lesson 1 Cheek Cell DNA Template Preparation (Lab Protocol)

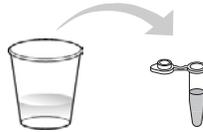
1. Each member of your team should have 1 screwcap tube containing 200 μ l InstaGene™ matrix, 1.5 ml micro test tube, and a cup containing 10 ml of 0.9% saline solution. Label one of each tube and a cup with your initials.



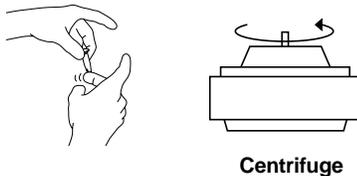
2. Do not throw away the saline after completing this step. Pour the saline from the cup into your mouth. Rinse vigorously for 30 seconds. Expel the saline back into the cup.



3. Set a P-1000 micropipet to 1,000 μ l and transfer 1 ml of your oral rinse into the micro test tube with your initials. If no P-1000 is available, carefully pour ~1 ml of your swished saline into the micro test tube (use the markings on the side of the micro test tube to estimate 1 ml).

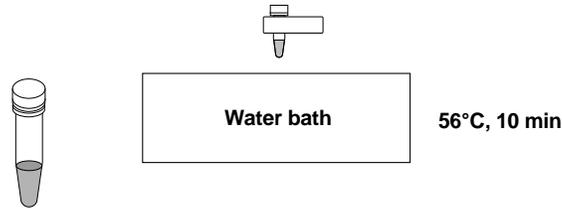


4. Spin your tube in a balanced centrifuge for 2 minutes at full speed. When the centrifuge has completely stopped, remove your tube. You should be able to see a pellet of whitish cells at the bottom of the tube. Ideally, the pellet should be about the size of a match head. If you can't see your pellet, or your pellet is too small, pour off the saline supernatant, add more of your saline rinse, and spin again.

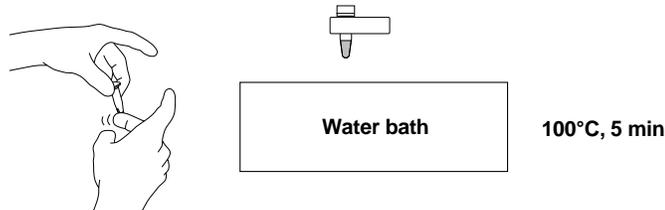


5. Pour off the supernatant and discard. Taking care not to lose your cell pellet, carefully blot your micro test tube on a tissue or paper towel. It's ok for a small amount of saline (~50 μ l, about the same size as your pellet) to remain in the bottom of the tube.
6. Resuspend the pellet thoroughly by vortexing or flicking the tubes until no cell clumps remain.

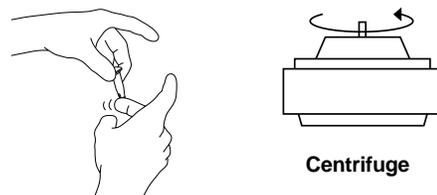
7. Using an adjustable volume micropipet set to 20 μ l, transfer your resuspended cells into the screwcap tube containing the InstaGene with your initials. You may need to use the pipet a few times to transfer all of your cells.
8. Screw the caps tightly on the tubes. Shake or vortex to mix the contents.
9. Place the tubes in the foam micro test-tube holder. When all members of your team have collected their samples, float the holder with tubes in a 56°C water bath for 10 minutes. At the halfway point (5 minutes), shake or vortex the tubes several times. Place the tubes back in the water bath for the remaining 5 minutes.



10. Remove the tubes from the water bath and shake them several times. Now float the holder with tubes in a 100°C water bath for 5 minutes.



11. Remove the tubes from the 100°C water bath and shake or vortex several times to resuspend the sample. Place the eight tubes in a balanced arrangement in a centrifuge. Pellet the matrix by spinning for 5 minutes at 6,000 x g (or 10 minutes at 2,000 x g).



12. Store your screwcap tube in the refrigerator until the next laboratory period, or proceed to step 2 of Lesson 2 if your teacher instructs you to do so.

Lesson 1 DNA Template Preparation

Focus Questions

1. Why is it necessary to chelate the metal ions from solution during the boiling/lysis step at 100°C? What would happen if you did not use a chelating agent such as the InstaGene matrix?
2. What is needed from the cells for PCR?
3. What structures must be broken to release the DNA from a cell?
4. Why do you think the DNA is stored cold with the InstaGene matrix after boiling the samples?

Lesson 2 PCR Amplification

It is estimated that there are 30,000–50,000 individual genes in the human genome. The true power of PCR is the ability to target and make millions of copies of (or **amplify**) a specific piece of DNA (or gene) out of a complete genome. In this activity, you will amplify a region within your chromosome 16.

The recipe for a PCR amplification of DNA contains a simple mixture of ingredients. To replicate a piece of DNA, the reaction mixture requires the following components:

1. DNA template — containing the intact sequence of DNA to be amplified
2. Individual deoxynucleotides (A, T, G, and C) — raw material of DNA
3. DNA polymerase — an enzyme that assembles the nucleotides into a new DNA chain
4. Magnesium ions — a cofactor (catalyst) required by DNA polymerase to create the DNA chain
5. Oligonucleotide primers — pieces of DNA complementary to the template that tell DNA polymerase exactly where to start making copies
6. Salt buffer — provides the optimum ionic environment and pH for the PCR reaction

The template DNA in this exercise is genomic DNA that was extracted from your cells. The complete master mix contains *Taq* DNA polymerase, deoxynucleotides, oligonucleotide primers, magnesium ions, and buffer. When all the other components are combined under the right conditions, a copy of the original double-stranded template DNA molecule is made — doubling the number of template strands. Each time this cycle is repeated, copies are made from copies and the number of template strands doubles — from 2 to 4 to 8 to 16 and so on — until after 20 cycles there are 1,048,576 exact copies of the target sequence.

PCR makes use of the same basic processes that cells use to duplicate their DNA.

1. **Complementary DNA strand hybridization**
2. **DNA strand synthesis via DNA polymerase**

The two DNA primers provided in this kit are designed to flank a DNA sequence within your genome and thus provide the exact start signal for the DNA polymerase to “zero in on” and begin synthesizing (replicating) copies of that target DNA. Complementary strand hybridization takes place when the two different **primers** anneal, or bind to each of their respective complementary base sequences on the template DNA.

The primers are two short single-stranded DNA molecules (23 bases long), one that is complementary to a portion of one strand of the template, and another that is complementary to a portion of the opposite strand. These primers anneal to the separated template strands and serve as starting points for DNA *Taq* replication by DNA polymerase.

Taq DNA polymerase extends the annealed primers by “reading” the template strand and synthesizing the complementary sequence. In this way, *Taq* polymerase replicates the two template DNA strands. This polymerase was isolated from a heat-stable bacterium (*Thermus aquaticus*) which in nature lives within high temperature steam vents such as those found in Yellowstone National Park.⁶ For this reason these enzymes have evolved to withstand high temperatures (94°C) and can be used in the PCR reaction.

PCR Step by Step

PCR amplification includes three main steps, a **denaturation step**, an **annealing step**, and an **extension step** (summarized in Figure 9). In denaturation, the reaction mixture is heated to 94°C for 1 minute, which results in the melting or separation of the double-stranded DNA template into two single stranded molecules. PCR amplification, DNA templates must be separated before the polymerase can generate a new copy. The high temperature required to melt the DNA strands normally would destroy the activity of most enzymes, but *Taq* polymerase is stable and active at high temperature.

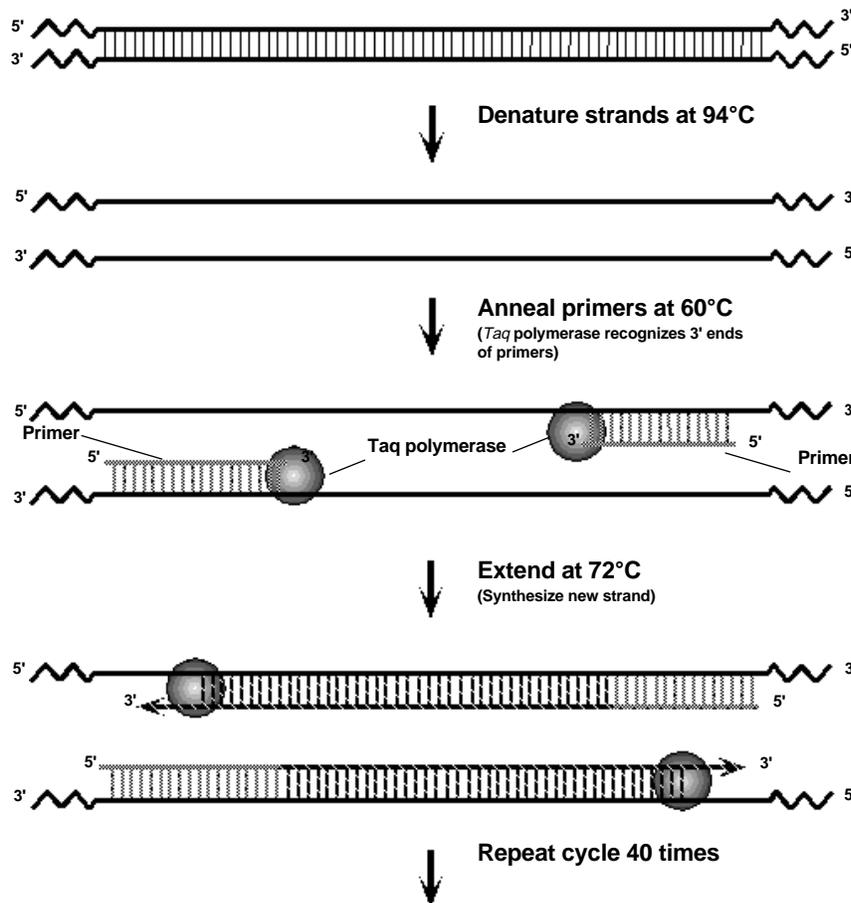


Fig. 9. A complete cycle of PCR.

During the annealing step, the oligonucleotide primers “anneal to” or find their complementary sequences on the two single-stranded template strands of DNA. In these annealed positions, they can act as primers for *Taq* DNA polymerase. They are called primers because they “prime” the synthesis of a new strand by providing a short sequence of double-stranded DNA for *Taq* polymerase to extend from and build a new complementary strand. Binding of the primers to their template sequences is also highly dependent on temperature. In this lab exercise, a 60°C annealing temperature is optimum for primer binding.

During the extension step, the job of *Taq* DNA polymerase is to add nucleotides (A, T, G, and C) one at a time to the primer to create a complementary copy of the DNA template. During

polymerization the reaction temperature is 72°C, the temperature that produces optimal *Taq* polymerase activity. The three steps of denaturation, annealing, and extension form one “cycle” of PCR. A complete PCR amplification undergoes 40 cycles.

The entire 40 cycle reaction is carried out in a test tube that has been placed into a thermal cycler. The thermal cycler contains an aluminum block that holds the samples and can be rapidly heated and cooled across broad temperature differences. The rapid heating and cooling of this thermal block is known as **temperature cycling** or **thermal cycling**.

Temperature Cycle = Denaturation Step (94°C) + Annealing Step (60°C) + Extension Step (72°C)

Lesson 2 PCR Amplification (Lab Protocol)

Workstation Checklist

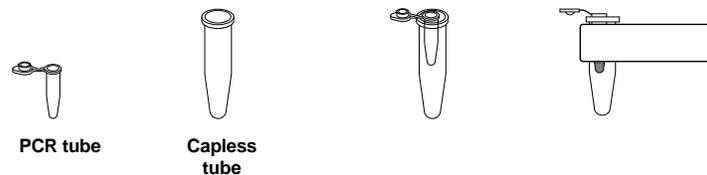
Materials and supplies that should be present at the workstations prior to beginning this lab are listed below.

Student Workstation	Quantity per Station	(✓)
PCR tubes	4	<input type="checkbox"/>
Micro test tubes, capless	4	<input type="checkbox"/>
Complete master mix (with primers) on ice	1 tube	<input type="checkbox"/>
P-20 micropipet	1	<input type="checkbox"/>
Pipet tips (filter type), 2–20 μ l	8	<input type="checkbox"/>
Ice bucket with ice	1	<input type="checkbox"/>
Foam micro test tube holders	2	<input type="checkbox"/>
Permanent marker	1	<input type="checkbox"/>
Waste containers	1	<input type="checkbox"/>
Copy of Quick Guide or protocol	1	<input type="checkbox"/>

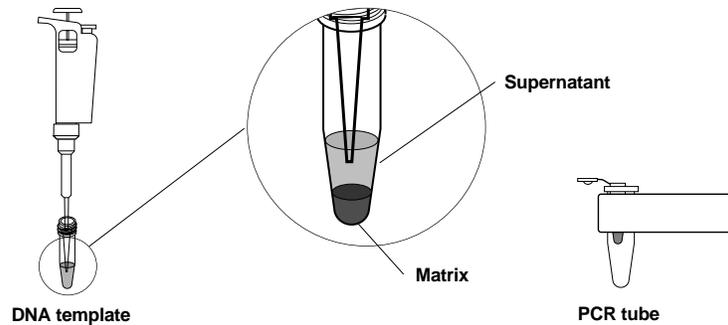
Instructor's (common) workstation	Quantity per Class	
Gel trays	1 per 2 stations	<input type="checkbox"/>
Molten agarose	40 ml per gel	<input type="checkbox"/>
Lab tape for gel trays	1 per station	<input type="checkbox"/>
Gene Cycler or MyCycler thermal cycler	1	<input type="checkbox"/>
Microcentrifuge or mini centrifuge	1 4	<input type="checkbox"/> <input type="checkbox"/>

Lesson 2 PCR Amplification (Lab Protocol)

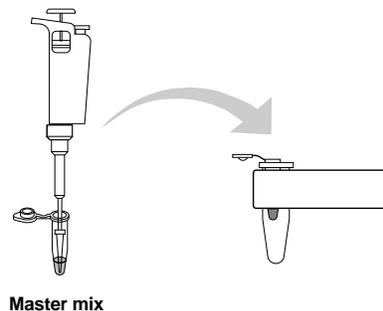
1. Obtain your screwcap tube that contains your genomic DNA template from the refrigerator. Centrifuge your tubes for 2 minutes at 6,000 x g or for 5 minutes at 2,000 x g in a centrifuge.
2. Each member of the team should obtain a PCR tube and capless micro test tube. Label each PCR tube on the side of the tube with your initials and place the PCR tube into the capless micro test tube as shown. Place the PCR tube in the foam micro test tube holder.



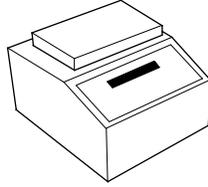
3. Transfer 20 μ l of your DNA template from the supernatant in your screwcap tube into the bottom of the PCR tube. **Do not transfer any of the matrix beads into the PCR reaction because they will inhibit the PCR reaction.**



4. Locate the tube of yellow PCR master mix (labeled "Master") in your ice bucket. Transfer 20 μ l of the master mix into your PCR tube. Mix by pipetting up and down 2–3 times. Cap the PCR tube tightly and keep it on ice until instructed to proceed to the next step. Avoid bubbles, especially in the bottom of the tubes.



5. Remove your PCR tube from the capless micro test tube and place the tube in the Gene Cyclor or MyCyclor thermal cyclor.



6. When all of the PCR samples are in the thermal cyclor, the teacher will begin the PCR reaction. The reaction will undergo 40 cycles of amplification, which will take approximately 3 hours.
7. If your teacher instructs you to do so, you will now pour your agarose gels (the gels may have been prepared ahead of time by the teacher).

Lesson 2 PCR Amplification

Focus Questions

1. Why is it necessary to have a primer on each side of the DNA segment to be amplified?
2. How did *Taq* DNA polymerase acquire its name?
3. Why are there nucleotides (A, T, G, and C) in the master mix? What are the other components of the master mix, and what are their functions?
4. Describe the three main steps of each cycle of PCR amplification and what reactions occur at each temperature.
5. Explain why the precise length target DNA sequence doesn't get amplified until the third cycle. You may need to use additional paper and a drawing to explain your answer.

Lesson 3 Gel Electrophoresis of Amplified PCR Samples and Staining of Agarose Gels

What Are You Looking At?

Before you analyze your PCR products, let's take a look at the target sequence being explored.

What Can Genes and DNA Tell Us?

It is estimated that the 23 pairs, or 46 **chromosomes**, of the human genome (23 chromosomes come from the mother and the other 23 come from the father) contain approximately 30,000–50,000 genes. Each chromosome contains a series of specific genes. The larger chromosomes contain more DNA, and therefore more genes, compared to the smaller chromosomes. Each of the homologous chromosome pairs contains similar genes.

Each gene holds the code for a particular protein. Interestingly, the 30,000–50,000 genes only comprise 5% of the total chromosomal DNA. The other 95% is noncoding DNA. This noncoding DNA is found not only between, but within genes, splitting them into segments. The exact function of the noncoding DNA is not known, although it is thought that noncoding DNA allows for the accumulation of mutations and variations in genomes.

When RNA is first transcribed from DNA, it contains both coding and noncoding sequences. While the RNA is still in the nucleus, the noncoding **introns** (**in = stay within** the nucleus) are removed from the RNA while the **exons** (**ex = exit the nucleus**) are spliced together to form the complete messenger RNA coding sequence for the protein (Figure 10). This process is called **RNA splicing** and is carried out by specialized enzymes called **spliceosomes**.

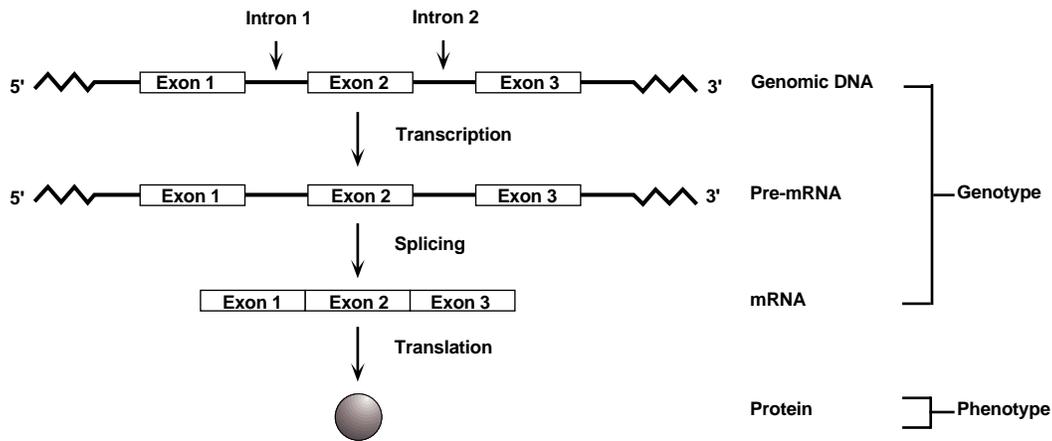


Fig. 10. Splicing of introns from genes.

Introns often vary in their size and sequence among individuals, while exons do not. This variation is thought to be the result of the differential accumulation of mutations in DNA throughout evolution. These mutations in our noncoding DNA are silently passed on to our descendants; we do not notice them because they do not affect our phenotypes. However, these differences in our DNA represent the molecular basis of DNA fingerprinting used in human identification and studies in population genetics.

The Target Sequence

The human genome contains small, repetitive DNA elements or sequences that have become randomly inserted into it over millions of years. One such repetitive element is called the “Alu sequence”⁷ (Figure 11). This is a DNA sequence about 300 base pairs long that is repeated almost 500,000 times throughout the human genome.⁸ The origin and function of these repeated sequences is not yet known.

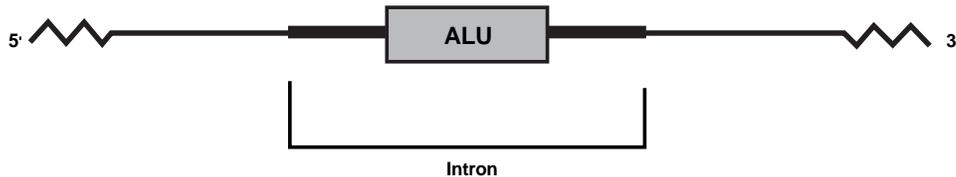


Fig. 11. Location of an Alu repetitive element within an intron.

Some of these Alu sequences have characteristics that make them very useful to geneticists. When present within introns of certain genes, they can either be associated with a disease or be used to estimate relatedness among individuals. In this exercise, analysis of a single Alu repeat is used to estimate its frequency in the population and as a simple measure of molecular genetic variation — **with no reference to disease or relatedness among individuals.**

In this laboratory activity you will look at an Alu element in the PV92 region of chromosome 16. This particular Alu element is **dimorphic**, meaning that the element is present in some individuals and not others. Some people have the insert in one copy of chromosome 16 (**one allele**), others may have the insert in both copies of chromosome 16 (**two alleles**), while some may not have the insert on either copy of the chromosome (Figure 12). The presence or absence of this insert can be detected using PCR followed by agarose gel electrophoresis.

Since you are amplifying a region of DNA contained within an intron, the region of DNA is never really used in your body. So if you don't have it, don't worry.

The primers in this kit are designed to bracket a sequence within the PV92 region that is 641 base pairs long if the intron does not contain the Alu insertion, or 941 base pairs long if Alu is present. This increase in size is due to the 300 base pair sequence contributed by the Alu insert.

When your PCR products are electrophoresed on an agarose gel, three distinct outcomes are possible. If both chromosomes contain Alu inserts, each amplified PCR product will be 941 base pairs long. On a gel they will migrate at the same speed so there will be one band that corresponds to 941 base pairs. If neither chromosome contains the insert, each amplified PCR product will be 641 base pairs and they will migrate as one band that corresponds to 641 base pairs. If there is an Alu insert on one chromosome but not the other, there will be one PCR product of 641 base pairs and one of 941 base pairs. The gel will reveal two bands for such a sample.

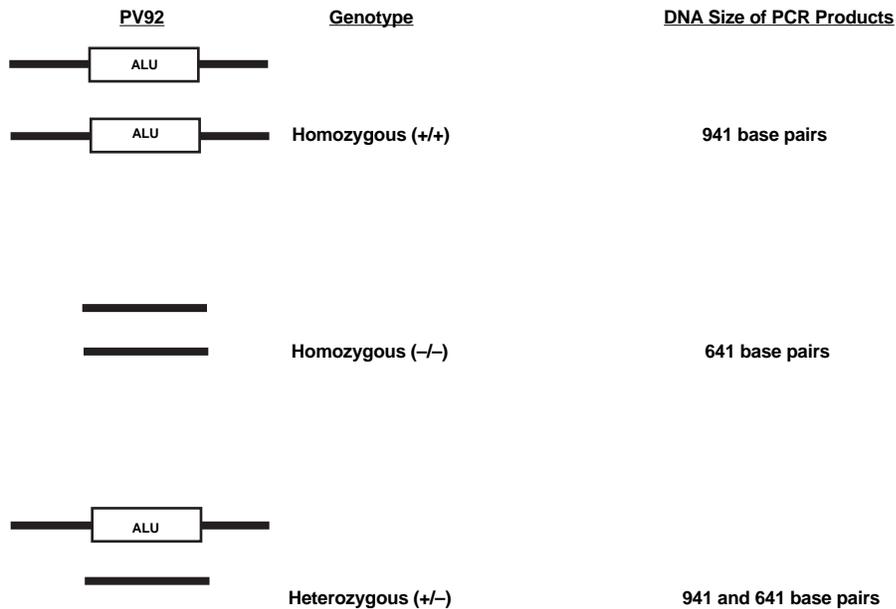


Fig. 12. The presence or absence of the Alu insert within the PV92 region of chromosome 16.

Electrophoresis separates DNA fragments according to their relative sizes. DNA fragments are loaded into an agarose gel slab, which is placed into a chamber filled with a conductive buffer solution. A direct current is passed between wire electrodes at each end of the chamber. DNA fragments are negatively charged, and when placed in an electric field will be drawn toward the positive pole and repelled by the negative pole. The matrix of the agarose gel acts as a molecular sieve through which smaller DNA fragments can move more easily than larger ones. Over a period of time, smaller fragments will travel farther than larger ones. Fragments of the same size stay together and migrate in what appears as a single “band” of DNA in the gel. In the sample gel below (Figure 13), PCR-amplified bands of 941 bp and 641 bp are separated based on their sizes.

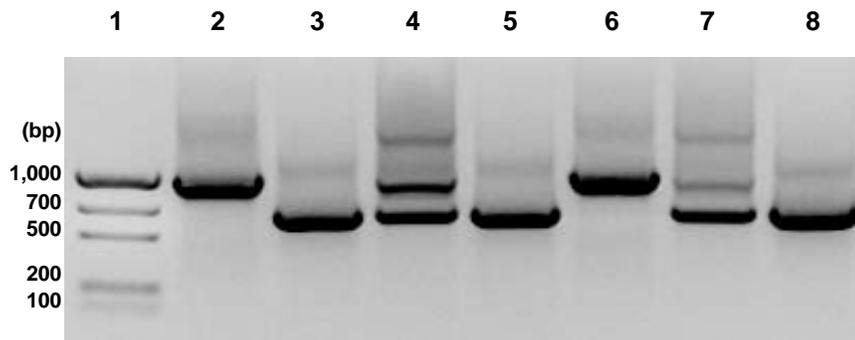


Fig. 13. Electrophoretic separation of DNA bands based on size. EZ Load DNA molecular mass ruler, which contains 1,000 bp, 700 bp, 500 bp, 200 bp, and 100 bp fragments (lane 1); two homozygous (+/+) individuals with 941 bp fragments (lanes 2, 6); three homozygous (-/-) individuals with 641 bp fragments (lanes 3, 5, and 8), and two heterozygous (+/-) individuals with 941 and 641 bp fragments (lanes 4 and 7).

Lesson 3 Gel Electrophoresis of Amplified PCR Samples and Staining of Agarose Gels (Lab Protocol)

Workstation Checklist

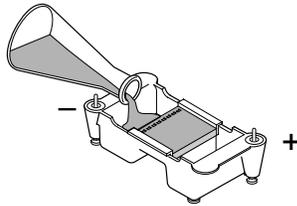
Materials and supplies that should be present at the workstations prior to beginning this lab are listed below.

Student workstation	Quantity per Station	(✓)
Agarose gel	1	<input type="checkbox"/>
Student PCR samples	1 per student	<input type="checkbox"/>
MMR (DNA standard)	1 tube	<input type="checkbox"/>
PV92 XC DNA loading dye	1 tube	<input type="checkbox"/>
P-20 micropipet	1	<input type="checkbox"/>
Pipet tips (filter type), 2–20 µl	12	<input type="checkbox"/>
Permanent marker	1	<input type="checkbox"/>
Foam micro test tube holder	1	<input type="checkbox"/>
Gel box and power supply	1	<input type="checkbox"/>
Fast Blast™ DNA stain, 1x or 100x solution	120 ml per 2 stations	<input type="checkbox"/>
Gel support film (optional)	1	<input type="checkbox"/>
Clear acetate sheets for tracing gels (optional)	1	<input type="checkbox"/>
Warm tap water for destaining gels (if performing quick staining protocol)	1.5–2 L per 2 stations	<input type="checkbox"/>
Large containers for destaining (if performing quick staining protocol)	1–3 per 2 stations	<input type="checkbox"/>
Copy of Quick Guide or protocol	1	<input type="checkbox"/>
Waste container	1	<input type="checkbox"/>
Instructor's workstation	Quantity per Class	
1x TAE electrophoresis buffer	275 ml per gel box	<input type="checkbox"/>
Amplified positive control samples (4 each)	12	<input type="checkbox"/>
PV92 homozygous (+/+)		
PV92 homozygous (-/-)		
PV92 heterozygous (+/-)		
Shaking platform (optional)*	1	<input type="checkbox"/>
Microcentrifuge	1	<input type="checkbox"/>
or mini centrifuge	4	<input type="checkbox"/>

* Strongly recommended.

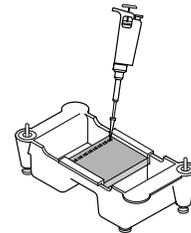
Lesson 3 Gel Electrophoresis of Amplified PCR Samples (Lab Protocol)

1. Remove your PCR samples from the thermal cycler and place in the micro test tube holder. If a centrifuge is available, place the PCR tubes in the capless micro test tubes and pulse-spin the tubes (~3 seconds at 2,000 x g) to bring the condensation that formed on the lids to the bottom of the tubes.
2. Add 10 μ l of PV92 XC loading dye to each PCR tube and mix gently.
3. Obtain an agarose gel (either the one you poured or one pre-poured by your teacher). Place the casting tray with the solidified gel in it, onto the platform in the gel box. The wells should be at the cathode (-) end of the box, where the black lead is connected. Very carefully remove the comb from the gel by pulling it straight up, slowly.
4. Pour ~275 ml of electrophoresis buffer into the electrophoresis chamber, until it just covers the wells.

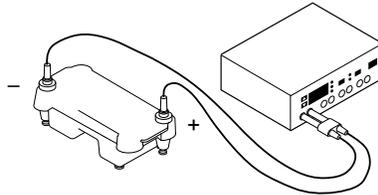


5. Using a clean tip for each sample, load the samples into the 8 wells of the gel in the following order:

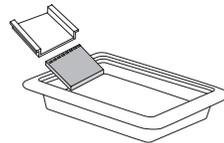
Lane	Sample	Load Volume
1	MMR (DNA standard)	10 μ l
2	Homozygous (+/+) control	10 μ l
3	Homozygous (-/-) control	10 μ l
4	Heterozygous (+/-) control	10 μ l
5	Student 1	20 μ l
6	Student 2	20 μ l
7	Student 3	20 μ l
8	Student 4	20 μ l



- Secure the lid on the gel box. The lid will attach to the base in only one orientation: red to red and black to black. Connect the electrical leads to the power supply.
- Turn on the power supply. Set it to 100 V and electrophorese the samples for 30 minutes.



- When electrophoresis is complete, turn off the power and remove the lid from the gel box. Carefully remove the gel tray and the gel from the gel box. Be careful, the gel is very slippery. Nudge the gel off the gel tray with your thumb and carefully slide it into your plastic staining tray.



Staining of Agarose Gels

The moment of truth has arrived. What is your genotype? Are you a homozygote or a heterozygote? To find out, you will have to stain your agarose gel. Since DNA is naturally colorless, it is not immediately visible in the gel. Unaided visual examination of gel after electrophoresis indicates only the positions of the loading dyes and not the positions of the DNA fragments. DNA fragments are visualized by staining the gel with a blue dye called Fast Blast DNA stain. The blue dye molecules are positively charged and have a high affinity for the DNA. These blue dye molecules strongly bind to the DNA fragments and allow DNA to become visible. These visible bands of DNA may then be traced, photographed, sketched, or retained as a permanently dried gel for analysis.

Directions for Using Fast Blast DNA Stain

Below are two protocols for using Fast Blast DNA stain in the classroom. Use protocol 1 for quick staining of gels to visualize DNA bands in 12–15 minutes, and protocol 2 for overnight staining. Depending on the amount of time available, your teacher will decide which protocol to use. Two student teams will stain the gels per staining tray (you may want to notch gel corners for identification). Mark staining trays with initials and class period before beginning this activity.

WARNING

Although Fast Blast DNA stain is nontoxic and noncarcinogenic, latex or vinyl gloves should be worn while handling the stain or stained gels to keep hands from becoming stained blue. Lab coats or other protective clothing should be worn to avoid staining clothes.

Protocol 2: Overnight Staining of Agarose Gels in 1x Fast Blast DNA Stain

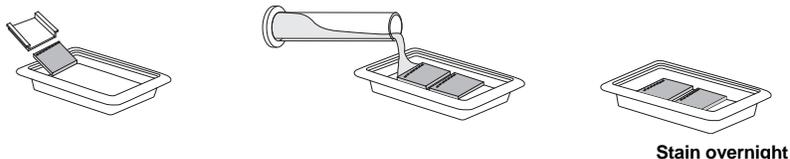
For overnight staining, Fast Blast DNA stain (500x) should be diluted to a 1x concentration. We recommend using 120 ml of 1x Fast Blast to stain two 7 x 7 cm or 7 x 10 cm agarose gels in individual staining trays provided in Bio-Rad's education kits. If alternative staining trays are used, add a sufficient volume of staining solution to completely submerge the gels.

Following DNA electrophoresis, agarose gels must be removed from their gel trays before being placed in the staining solution. This is easily accomplished by holding the base of the gel tray in one hand and gently pushing out the gel with the thumb of the other hand. Because the gel is fragile, special attention must be given when handling it.

1. Mark staining trays with your initials and class period. You will stain two gels per tray.

2. Stain gels (overnight)*

Pour 1x stain into a gel staining tray. Remove each gel from the gel tray and carefully slide it into the staining tray containing the stain. If necessary, add more 1x staining solution to completely submerge the gels. Place the staining tray on a rocking platform and agitate overnight. If no rocking platform is available, agitate the gels staining tray a few times during the staining period. You should begin to see DNA bands after 2 hours, but at least 8 hours of staining is recommended for complete visibility of stained bands.



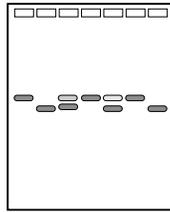
3. Analyze results

No destaining is required after staining with 1x Fast Blast. The gels can be analyzed immediately after staining.

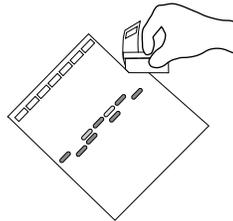
- a. Place your gel on a light background and record your results by making a diagram as follows. Place a clear sheet of plastic sheet or acetate over the gel. With a permanent marker, trace the wells and band patterns onto the plastic sheet to make a replica picture of your gel. Remove the plastic sheet for later analysis. Alternatively, gels can be photocopied on a yellow piece of transparent film for optimal contrast.

* Shake the gels gently and intermittently during overnight staining in 1x Fast Blast DNA stain; small DNA fragments tend to diffuse without shaking.

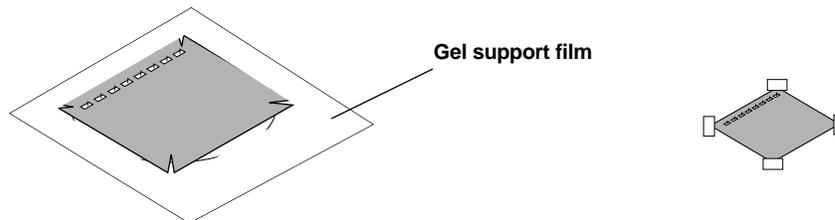
- b. With the help of your instructor, determine whether you are homozygous or heterozygous for the Alu insertion. First look at the control samples and note the migration patterns of the homozygous +/+, the homozygous -/-, and the heterozygous +/- samples (also refer to the example on page 59). You may notice that in the heterozygous sample the smaller 641 base pair band is more intense than the larger 941 bp band. This difference is due to the fact that the smaller fragment is amplified more efficiently than the larger fragment. Copies of the shorter fragment can be made at a faster rate than the bigger fragment, so more copies of the shorter fragment are created per cycle. Refer to pages 69–72 for more information on how to analyze your data.



- c. Dry the agarose gel as a permanent record of the experiment.
- Trim away any unloaded lanes with a knife or razor blade. Cut your gel from top to bottom to remove the lanes that you did not load samples into.



- Place the gel directly upon the hydrophilic side of a piece of gel support film. (Water will form beads on the hydrophobic side of a piece of gel support film.) Center the gel on the film on a paper towel and let the gel dry in a well-ventilated area, making sure to avoid direct exposure to light. As the gel dries it will bond to the film but will not shrink. If left undisturbed on the support film, the gel will dry completely at room temperature after 2–3 days. The result will be a flat, transparent, and durable record for the experiment. Tape the dried gel into your laboratory notebook.



Note: Avoid extended exposure of dried gels to direct light to prevent band fading. However, DNA bands will reappear if the dried gels are stored in the dark for 2–3 weeks after fading.

Lesson 3 Gel Electrophoresis of Amplified PCR Samples

Focus Questions

1. Explain the difference between an intron and an exon.
2. Why do the two possible PCR products differ in size by 300 base pairs?
3. Explain how agarose electrophoresis separates DNA fragments. Why does a smaller DNA fragment move faster than a larger one?
4. What kind of controls are run in this experiment? Why are they important? Could others be used?

Lesson 4 Analysis and Interpretation of Results

If the overnight staining protocol was used to stain the gels, record your results and dry your gels as described earlier.

Analysis

Compare your sample lanes with the control lanes using the DNA size marker as a reference. Mark the location and size of your fragment or fragments. By comparing your DNA migration pattern to the controls, determine whether you are homozygous +/+, homozygous -/-, or heterozygous +/- . If your sample lane is blank, discuss the possible reasons with your classmates and teacher.

Remember that the interpretation of this gel allows you to determine your genetic makeup only at the locus (chromosomal location), being studied. There are three possible genotypes for the Alu insert at the location you have amplified. For a class, determine the number of individuals of each genotype: homozygous +/+, homozygous -/-, and heterozygous +/- . Tally the class results in the table on page 70.

A major factor affecting the reliability of DNA fingerprinting evidence in forensics is population genetics and genetic statistics. In humans, there are hundreds of loci, or DNA segments, like Alu, that can be selected and used for fingerprinting analysis. Depending on demographic factors such as ethnicity and geographic isolation, some populations show much less variation in particular DNA segments than others. A lower degree of variation will increase the odds of more than one individual having the same sequence. If 33% (1 out of three individuals) of a given population has the same fingerprinting pattern for a certain DNA segment, then little information will be obtained from using that segment alone to identify an individual. In such a case, a positive result would only identify a person with 33% accuracy.

In analyzing how incriminating the DNA evidence is, one needs to ask the question: Statistically, how many people in a population have the same DNA pattern as that taken from a crime scene: 1 in 1,000,000? 1 in 10,000? 1 in 10?

For a DNA fingerprint to identify a suspect in a criminal case or a father in a paternity suit, accurate identification required not a 1 out of 3 (1/3) chance of a match in a population, but closer to a 1 in 10 million (1/10⁷) chance of a match. The frequency of a particular DNA pattern turning up in a population drastically decreases when multiple DNA segments are selected and amplified, rather than just one segment. For DNA fingerprinting to be admissible as evidence in court, it must analyze 30 to 40 different DNA segments on multiple chromosomes from the same person.

The Alu insert you have fingerprinted in this exercise has been used to study the migration patterns of human populations over time.⁸ The data from these studies have been published, and your class samples can be compared to the data collected from much larger populations.

Appendix A

Review of Molecular Biology

This section provides an overview and concepts with which students should be familiar in order to get the most out of this lab. Please also refer to the Glossary Section (Appendix B) for definitions of molecular biology terms.

Any living organism functions based on the complicated interactions among nucleic acids, proteins, lipids (fat), and carbohydrates. In nearly all cases, certain proteins, termed enzymes, control the almost infinite number of interactions and life processes in living creatures. Think of enzymes and proteins as all the different people on earth. Each person performs a different role, function, or job on this planet, and although people are not the actual physical make-up of buildings, documents, food, and roads, it is the people that make these buildings and roads, and write the documents, and plant and nurture the crops. In the same way, enzymes and proteins do not comprise bones, lipids, sex hormones, and sugars, but enzymes control these structures, their interactions, and processes.

Because proteins and enzymes ultimately play such a critical role in the life process, scientists have spent many lifetimes studying proteins in an attempt to understand how they work and how they can be controlled. With a complete understanding, we could cure, prevent, and overcome many diseases and physical handicaps as well as explain exactly how and why organisms exist, propagate, and die. However, the complete answers do not lie solely in the knowledge of how enzymes function; we must learn how they are made. Before we can control enzymes, we must understand where they come from and what is the basis of the molecular information that encodes proteins. That answer lies within our **genetic code**.

Each living organism has its own blueprint for life. This blueprint defines how an organism will look and function (using enzymes as a means to form the appearance and control the functions). The blueprint codes for all the different enzymes. With amazing precision, this blueprint gets passed on from generation to generation of each species.

The transfer of this blueprint from generation to generation is called **heredity**. The blueprint for any organism is called its **genome**. The hereditary code is encrypted within the sequence of the DNA molecules that make up the genome. The molecule that constitutes the genome and thus the hereditary code is DNA (**deoxyribonucleic acid**).

The genome consists of very long DNA/protein complexes called **chromosomes**. Prokaryotes, organisms lacking a true nucleus, have only one chromosome. All other species, eukaryotes, have a defined cell nucleus that contains multiple chromosomes. The nucleus is a defined, membrane-enclosed region of the cell that contains the chromosomes. The number of chromosomes varies with the organism — from 2 or 3 in some yeasts to up to 100 or so in some fish. Humans have 46.

In most cases, chromosomes come in nearly identical pairs (one member of the chromosome pair from each parent). In general, the members of a pair differ in small details from each other, since they come from different parents, but are otherwise identical or **homologous**. Cells with homologous pairs of chromosomes are called diploid. Nearly all cells of an organism are diploid. Cells that have only one chromosome of each pair are called haploid. All sperm and ova are haploid.

The process of forming sperm and ova is called **meiosis**. **Meiosis** starts with a diploid cell that divides into two haploid cells. When a sperm fertilizes an ovum, the two nuclei fuse, and thus the new nucleus contains pairs of each chromosome, one partner from each parent. The result is called a diploid zygote.

All cells of diploid organisms duplicate chromosomal pairs when they divide (except when sperm and ova are formed), so that all body cells (called somatic cells) of an organism are diploid. The process of cell division in which the chromosomes are duplicated and each daughter cell gets pairs of chromosomes is called **mitosis**. It is through the processes of mitosis and meiosis that the hereditary code is passed from cell to cell and generation to generation. Now that we know where the code is and how that code is passed on, we need to know how the code produces the enzymes that control life. The actual DNA code for a protein is contained within a segment of a chromosome called a gene. In nearly all cases, diploid organisms will have the same gene on a specific chromosome pair. Each gene on a particular chromosome of a specific chromosome pair is also called an **allele**.

To clarify, a gene encodes a particular protein that performs a particular function. An allele is a specific version of a gene on a particular chromosome. Thus, there are genes for hair color and there is an allele for the hair color gene on each chromosome pair. The gene or allele's DNA code can also be called the genotype.

When the protein is made from this code and performs its function, the physical trait or result that is seen is called the **phenotype**. In many cases the two alleles on the specific chromosome pair coding for a protein differ slightly in their respective DNA code (genotype). Any slight difference in code between the two alleles can result in two different proteins, which, although intended to perform basically the same function, may carry out that function slightly differently, causing different results and thus different phenotypes.

Therefore, it is not only the various combinations of chromosomes a parent contributes to each offspring, but also the various combinations of alleles and how each of the enzymes coded from the alleles work together that decide how we look and allow us to function. The various combinations are nearly infinite and that is why we are all different. The study of genotypes and phenotypes is often referred to as **Mendelian genetics** (after Mendel, the individual who pioneered the study of heredity and genetics).

DNA: What Is It?

A DNA molecule is a long polymer consisting of four different components called **bases**. The four bases are also called **nucleotides**. It is the various combinations of these four bases or nucleotides that create a unique DNA code or sequence (also genotype, gene, and allele). Nucleotides are comprised of three different components:

- **Nitrogen base**
- **Deoxyribose sugar**
- **Phosphate group**

Each nucleotide contains the same ribose sugar and the phosphate group. What makes each nucleotide unique is its nitrogen base. There are four nitrogen bases:

Adenine (A)

Thymine (T)

Guanine (G)

Cytosine (C)

A DNA nucleotide chain is created by the connection of the phosphate group to the ribose sugar of the next nucleotide. This connection creates the “backbone” of the DNA molecule.

To designate the different ends of this single-stranded chain, we use some typical biochemistry terminology, in which the carbons on any sugar are numbered. The sugar of a nucleotide contains 5 carbons. The phosphate group (PO_4) of a given nucleotide is connected to the 5' carbon of the sugar. A hydroxyl group (OH) is attached to the 3' carbon of the sugar, and this 3' OH group connects to the phosphate group of the next nucleotide in the chain.

Thus, the end of a single-strand DNA molecule that has a free phosphate group (i.e., not attached to another nucleotide) is called the 5' end, and the end of the DNA molecule (with no subsequent nucleotide attached) is called the 3' end (see Figures 14 and 15).

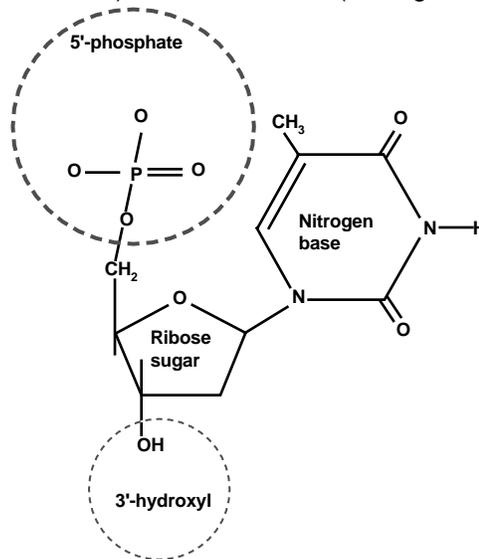


Fig. 14. Structure of one nucleotide of deoxyribonucleic acid.

It has become standard that a single-stranded DNA molecule is written with the 5' end on the left and the 3' end on the right. Therefore, a single-stranded DNA chain's sequence is represented from left to right, starting on the left with the 5' nucleotide and moving to the right until the 3' nucleotide is last. Most DNA sequences are read 5' to 3'.

However, the long DNA molecules or chains that comprise the chromosomes are not single-stranded molecules. From X-ray crystallography patterns of DNA, and some imaginative molecular model building, Watson and Crick deduced that DNA is in fact a **double-stranded** molecule with the two single strands of DNA held together by **hydrogen bonds** between the nitrogen bases (A, T, G, and C). This double-stranded molecule is often

called a duplex (Figure 15). There are several important properties of double-stranded DNA molecules.

- Chromosomal (also called genomic) DNA is double-stranded.
- The overall structure is that of a helix with two strands intertwined.
- The structure can be viewed as a twisted ladder.
- The phosphate-deoxyribose backbones are the sides of the ladder.
- The nitrogen bases (A, T, G, and C) hydrogen bonded to each other are the rungs.
- Only the nitrogen bases A and T and C and G can form hydrogen bonds to each other. When A binds to T or C binds to G this is considered **base pairing**. Neither C and T, nor A and G form hydrogen bonds.
- The two strands are antiparallel; that is, the strands are oriented in opposite directions. This means that the ladder runs 5' to 3' in one direction for one strand and 5' to 3' in the opposite direction for the other strand.

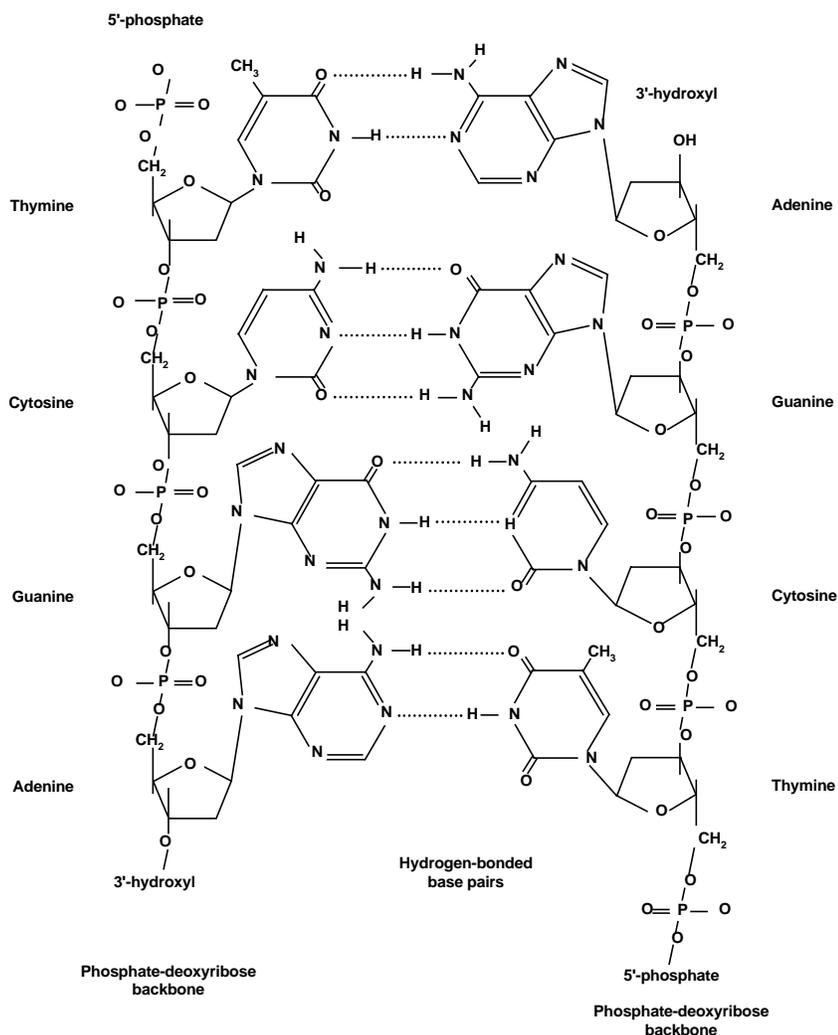


Fig. 15. Molecular structure of a portion of a double-stranded DNA molecule.

DNA Structure Conclusions

- Because A only binds to T, and G only binds to C, the two strands will have exactly the opposite, or complementary, sequence running in opposite directions (one strand 5' to 3', the other 3' to 5').
- These two complementary strands anneal or hybridize to each other through hydrogen bonds between the bases.
- A new strand of DNA can be synthesized using its complementary strand as the template for new synthesis.
- Each strand carries the potential to deliver and code for information.

The length of any double-stranded DNA molecule is given in terms of base pairs (bp). If a DNA strand contains over a thousand base pairs, the unit of measure is kilobases (1 kb = 1,000 bp). If there are over one million base pairs in a strand the unit of measure is megabases (1 Mb = 1,000 kb).

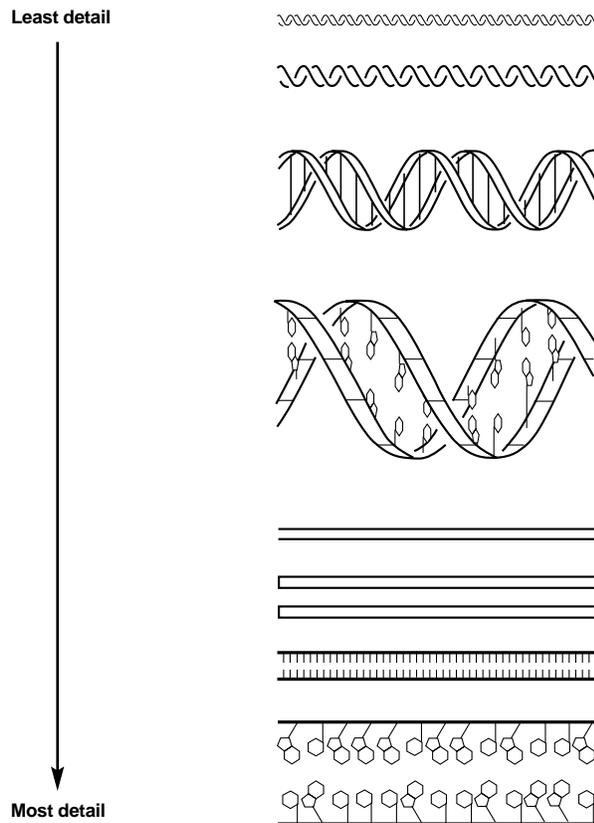


Fig. 16. DNA (deoxyribonucleic acid) — A long chainlike molecule that stores genetic information. DNA is graphically represented in a number of different ways, depending on the amount of detail desired.

DNA Replication — Strand Synthesis

New strands are synthesized by enzymes called **DNA polymerases**. New strands are always synthesized in the 5' to 3' direction. For a new single strand of DNA to be synthesized, another single strand is necessary. The single strand of DNA that will be used to synthesize its complementary strand is called the **template strand**.

However, in order for DNA polymerase to start synthesizing a new complementary strand, a short stretch of nucleotides (approximately 20 base pairs long) called an oligonucleotide primer must be present for the polymerase to start synthesis. This primer is a short stand of nucleotides complementary to the template where the researcher wants synthesis to begin. The primer must have a free 3' hydroxyl group (OH) for DNA polymerase to attach the 5' phosphate group of the next nucleotide.

The DNA polymerase grabs free (single) nucleotides from the surrounding environment and joins the 5' phosphate of the new nucleotide to the 3' hydroxyl group (OH) of the new complementary strand. This 5' to 3' joining process creates the backbone of the new DNA strand.

The newly synthesized strand maintains its complementarity with the template strand because the DNA polymerase only joins two nucleotides during new strand synthesis if the new nucleotide has its complement on the template strand. For example, the DNA polymerase will only join a G to the 3' end of the newly synthesized strand if there is the C counterpart on the template strand to form a hydrogen bond. Guanine will not be joined to the new strand if adenine, thymine, or guanine is the opposite nucleotide on the template strand.

DNA polymerase and strand synthesis allow DNA to replicate during mitosis. Both new DNA strands are synthesized simultaneously from the two original DNA template strands during mitotic DNA replication.

As you can see, DNA, RNA, and proteins are closely tied to each other. Thus, you can realize why researchers today, in an attempt to understand the mechanisms behind the various life processes, must study the nucleic acids as well as the proteins to get complete answers about the flow of information carried in the genetic code. In the last 20 years, many gains in the areas of nucleic acid techniques have finally allowed researchers the means to study the roles of nucleic acids in life processes.

Individual discoveries by many scientists have contributed the pieces that have begun to solve one of the most mysterious puzzles of life — understanding the hereditary code. In 1985, enough pieces of the puzzle were in place for a major breakthrough to occur. This understanding of how the necessary molecular components interact to faithfully replicate DNA within living cells led to the development of a technique for creating DNA in a test tube. This technique is called the **polymerase chain reaction**, or **PCR**.

Appendix B

Glossary of Terms

Aliquot	The division of a quantity of material into smaller, equal parts
Allele	A variation of a gene on a particular chromosome
Alu	A small piece of repetitive DNA that contains the <i>AluI</i> restriction enzyme site, from which the sequence obtained its name
Annealing	Binding of oligonucleotide primers to complementary sequences on the template DNA strands
Biotechnology	The manipulation of organisms (microbes, plants or animals) DNA to help solve human problems
Chelate	To bind metal ions in solution. An example of a common chelating agent is EDTA or ethylenediamine tetraacetic acid
Cofactors	Ions or small molecules needed by an enzyme to function properly. For example, <i>Taq</i> DNA polymerase needs Mg^{2+} in order to function properly. Mg^{2+} would therefore be considered a cofactor
Denature	The process of melting apart two complementary DNA strands. In vivo denaturation is accomplished by enzymes; in the PCR reaction denaturation is accomplished by heat
DNases	Digestive enzymes that degrade DNA
dNTPs	Commonly used abbreviation for all four deoxynucleotide triphosphates (dATP, dTTP, dGTP, dCTP) used in synthesizing DNA
Ethidium bromide	A fluorescent dye molecule that intercalates between DNA base pairs and fluoresces when exposed to ultraviolet light
Eukaryotes	Organisms that are made up of cells containing a membrane-bound nucleus that contains the genetic material (DNA)
Exon	The region of a transcribed messenger RNA molecule that gets spliced together and leaves the nucleus for translation into protein sequence
Extension	This refers to the process of <i>Taq</i> polymerase adding dNTPs (deoxynucleotide triphosphates — dATP, dTTP, dCTP, or dGTP) onto the ends of the oligonucleotide primers. Extension follows the base pairing rule and proceeds in the 5' to 3' direction
Genome	The sequence of DNA molecules within the nucleus that codes for all proteins for a given species. Each segment of DNA that encodes a given protein is called a gene. The information contained in the genome constitutes the organism's hereditary code
Genomic DNA	The sum total of the DNA that is found within the nucleus of a cell
Genotype	The combination of alleles carried by an individual

Appendix C

PCR Amplification and Sterile Technique

PCR is a powerful and sensitive technique that enables researchers to produce large quantities of DNA from very small amounts of starting material. Because of this sensitivity, contamination of PCR reactions with unwanted, extraneous DNA is always a possibility. Therefore, utmost care must be taken to prevent cross-contamination of samples. Steps to be taken to prevent contamination and failed experiments include:

1. **Filter-type pipet tips.** The end of the barrel of micropipets can easily become contaminated with DNA molecules that are aerosolized. Pipet tips that contain a filter at the end can prevent aerosol contamination from micropipets. DNA molecules within the micropipet can not pass through the filter and can not contaminate PCR reactions. Xcluda aerosol barrier pipet tips (catalog #211-2006EDU and 211-2016EDU) are ideal pipet tips to use in PCR reactions.
2. **Aliquot reagents.** Sharing of reagents and multiple pipettings into the same reagent tube will most likely introduce contaminants into your PCR reactions. When at all possible, aliquot reagents into small portions for each team, or if possible, for each student. If an aliquotted reagent tube does become contaminated, then only a minimal number of PCR reactions will become contaminated and fail.
3. **Change pipet tips.** Always change pipet tips when pipeting from a reagent for the first time. If a pipet tip is used repeatedly, contaminating DNA molecules on the tip will be passed into other solutions, resulting in contaminated PCR reactions. If you are at all unsure whether your pipet tip is clean, use the safe rule of thumb and discard the tip and get a new one. The price of a few extra tips is a lot smaller than the price of failed reactions.
4. **Use good sterile technique.** When opening, aliquotting, or pipetting reagents, leave the tube open for as little time as possible. Tubes that are open and exposed to the air can easily become contaminated by DNA molecules that are aerosolized or are present from your mouth, breath, etc. Pipet from reagent tubes efficiently, and close them when you are finished pipetting. Also, try not to pick tubes up by the rim or cap as you can easily introduce contaminating DNA molecules from your fingertips.

Hardy-Weinberg equilibrium	The conditions that enable a population to maintain its genetic frequencies; These conditions are: large population, random mating, no immigration or emigration, no mutations, and no natural selection
Homologous chromosomes	A pair of complementary chromosomes that contain the same genetic sequences, or genes, with one chromosome inherited from the mother and one chromosome inherited from the father
InstaGene matrix	Microscopic beads that bind divalent cations in solution; The binding or sequestering of divalent cations prevents their availability to enzymes that can degrade the DNA template
Intron	The region of a transcribed messenger RNA that is spliced out of the mRNA and is not translated into protein sequence
Lysis	The process of rupturing a cell to release its constituents. In this exercise, human cheek cells are lysed to release genomic DNA for PCR reactions
Master mix	The main solution of a PCR reaction which contains all of the necessary components (dNTPs, primer, buffer, salts, polymerase, magnesium) of the reaction except the template DNA
Messenger RNA	A type of RNA that is synthesized from the genetic material (DNA) and that attaches to ribosomes and is translated into protein
Molecular biology	The study of genes and the molecular details that regulate the flow of genetic information from DNA to RNA to proteins, and from generation to generation
Nucleotides	The fundamental unit of DNA or RNA; they consist of a sugar (deoxyribose or ribose), phosphate, and nitrogenous base (adenine, thymine, cytosine, or guanine, with uracil in place of thymine in RNA)
PCR	Polymerase chain reaction. The process of amplifying or synthesizing DNA within a test tube
Primer	A short sequence of nucleotides (usually 16–24 bases in length) that recognizes a particular sequence of nucleotides on the target DNA sequence; primers for the polymerase chain reaction are usually synthesized in a laboratory
Reagents	Materials needed to conduct an experiment; they are usually solutions or mixtures of various solutions
Taq DNA polymerase	Thermostable DNA polymerase that was isolated from the thermophilic bacterium <i>Thermus aquaticus</i> . This DNA polymerase is commonly used in PCR reactions
Template	The strand of DNA that contains the target sequences of the oligonucleotide primers and that will be copied into its complementary strand

Appendix F

References

1. Practice of the polymerase chain reaction (PCR) process requires a license. The Gene Cyclor and MyCyclor thermal cyclor are Authorized Thermal Cyclors may be used with PCR licenses available from Applied Biosystems. Their use with Authorized Reagents also provides a limited PCR license in accordance with the label rights accompanying such reagents. Some applications may also require licenses from other third parties.
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Appendix G Gel Loading Template

Use the template to enter students' names into the appropriate gel and well numbers.

	Controls			Student samples				
MMR	+/+	-/-	+/-	S1	S2	S3	S4	
	1	2	3	4	5	6	7	8
	<input type="text"/>							

MMR = molecular mass ruler (DNA standard)